

# FR209602 and Related Compounds, Novel Antifungal Lipopeptides from *Coleophoma crateriformis* No. 738

## II. *In Vitro* and *In Vivo* Antifungal Activity

Ryuichi Kanasaki, Fumie Abe, Shigetada Furukawa, Koji Yoshikawa, Akihiko Fujie, Motohiro Hino, Seiji Hashimoto, Yasuhiro Hori

Received: September 7, 2005 / Accepted: February 24, 2006

© Japan Antibiotics Research Association

**Abstract** The biological activities of the novel echinocandin-like lipopeptides, FR209602, FR209603 and FR209604, were evaluated. These compounds showed antifungal activity against *Candida albicans* and *Aspergillus fumigatus* attributed to inhibition of 1,3- $\beta$ -glucan synthesis. The minimum effective concentrations of these compounds against *C. albicans* and *A. fumigatus* ranged from 0.02 to 0.04  $\mu\text{g/ml}$  by microbroth dilution assay, and the  $\text{IC}_{50}$  values on *C. albicans* 1,3- $\beta$ -glucan synthase were 0.49, 0.64 and 0.72  $\mu\text{g/ml}$ , respectively. FR209602 and FR209603 showed good efficacy by subcutaneous injection against *C. albicans* in a murine systemic infection model, with  $\text{ED}_{50}$  values of 2.0 and 1.9 mg/kg, respectively.

**Keywords** FR209602, FR209603, FR209604, antifungal, 1,3- $\beta$ -glucan synthesis

### Introduction

Life-threatening infections caused by *Candida albicans* and *Aspergillus fumigatus* have been increasing in prevalence [1]. Antifungal therapies, however, are currently limited to a small number of compounds. Toxicity is an issue with treatments based on amphotericin B [2] and resistance is

R. Kanasaki (Corresponding authors), F. Abe, S. Furukawa, K. Yoshikawa, A. Fujie, M. Hino, S. Hashimoto, Y. Hori: Fermentation Research Laboratories, Fujisawa Pharmaceutical Co., Ltd., 5-2-3 Tokodai, Tsukuba, Ibaraki 300-2698, Japan, E-mail: ryuichi.kanasaki@jp.astellas.com

beginning to emerge as a problem with the safer but fungistatic azoles [3]. Therefore, new, safe and effective therapeutic agents are highly desirable for the treatment of infections caused by *Candida albicans* and *A. fumigatus*.

In the previous paper [4], we described the novel antifungal lipopeptides, FR209602, FR209603 and FR209604, produced by *Coleophoma crateriformis* No. 738. These compounds have potent antifungal activity against *Candida albicans* and *A. fumigatus* attributed to inhibition of 1,3- $\beta$ -glucan synthesis. Fungal cell wall components, especially 1,3- $\beta$ -glucan, are promising targets for antifungals, because this is essential to fungi and absent from mammalian cells [5]. In recent years, the new antifungal, micafungin [6–8] and caspofungin [9], which belong to the echinocandin-like family of lipopeptides, have been successfully launched as antifungal drugs.

In this paper, we describe the *in vitro* and *in vivo* antifungal activities of the newly isolated lipopeptides, FR209602, FR209603 and FR209604.

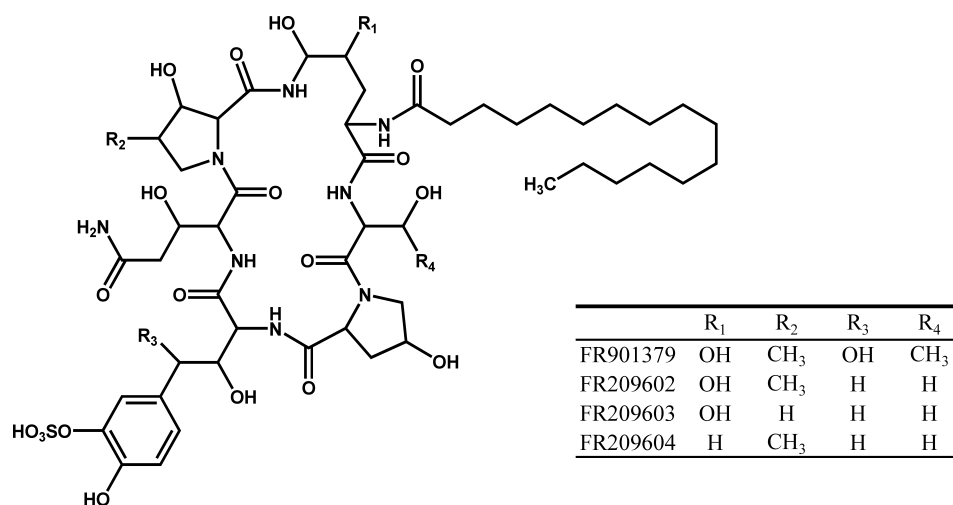
### Materials and Methods

#### Compound

FR901379 was isolated from the culture broth of *Coleophoma empetri* F-11899 which is a strain in the Fujisawa culture collection [10, 11].

#### *In Vitro* Antifungal Activity

Antifungal activity was measured by the micro-broth dilution method in 96-well culture plates employing yeast nitrogen base - dextrose (YNBD) medium. *Candida albicans*



**Fig. 1** Structures of FR901379, FR209602, FR209603 and FR209604.

FP633, a clinical isolate in the Fujisawa culture collection, was incubated in yeast-maltose (YM) broth for 20 hours at 37°C at the standing condition. The culture of *Cryptococcus neoformans* YC203 was grown in YM broth medium for 20 hours at 30°C with shaking. The cell suspension was prepared by washing the cultured cells with sterile saline. *A. fumigatus* FP1305, a strain from the Fujisawa culture collection, was cultured on YM agar slant for 7 days. The spores were harvested in sterile saline, and filtered through gauze. Finally, the fungal cells or spores were resuspended in yeast nitrogen base - dextrose medium for inoculation. Test compounds were diluted serially two fold with YNBD. The test microorganism was inoculated to each well to yield  $1 \times 10^4$  cfu/well in 100  $\mu$ l. The plates were incubated for 20 hours at 37°C (*Candida albicans* FP633 and *A. fumigatus* FP1305) or 48 hours at 37°C (*Cryptococcus neoformans* YC203). Minimum effective concentration (MEC) was determined by microscopic observation.

#### Membrane Fraction Preparation of *Candida albicans*

Membrane fraction preparation from *Candida albicans* was conducted according to the method described by Sawistowska-Schröder *et al.* [12] with some modifications. Briefly, yeast cells of *Candida albicans* FP633 were grown to logarithmic phase (absorbance at 660 nm; 0.42) in YNBD medium at 30°C with shaking. The cells were harvested by centrifugation, washed and suspended in ice-cold buffer A (50 mM Tris-HCl (pH 7.5), 1.0 mM  $\beta$ -mercaptoethanol, 1.0 M sucrose and 25  $\mu$ M GTP). The cells were sonicated with 0.4 mm i.d. glass beads using a sonicator. The glass beads were then washed with ice-cold buffer B (buffer A without sucrose), and the cell debris were removed by centrifugation. The supernatant fluids were centrifuged at  $100,000 \times g$  for 45 minutes at 4°C. The

pellet was washed with buffer B, resuspended in buffer C (buffer B - glycerol, 2 : 1) at 10 mg protein/ml and stored at -80°C as a source of particulate enzyme for glucan synthase assay.

#### Glucan Synthase Assay

The assay procedure for *Candida albicans* glucan synthase was conducted according to the method described by Sawistowska-Schröder *et al.* [12] with some modifications. Two and a half  $\mu$ l of test compound solution or vehicle was incubated with 25  $\mu$ l of reaction mixture (50 mM Tris-HCl (pH 8.0), 0.8% BSA, 0.1 mM GTP, 0.1% CHAPS, 0.05% Tween80 and the particulate enzyme (40  $\mu$ g protein)) for 15 minutes at room temperature. After incubation, 25  $\mu$ l of UDP-[U-<sup>14</sup>C]glucose (0.35  $\mu$ Ci/ml, 1.0 mM) was added to the reaction vessel to react for 60 minutes at room temperature. The reaction was terminated by addition of 100  $\mu$ l ice-cold 10% trichloroacetic acid (TCA) and allowed to stand on ice. The resultant precipitate was dissolved with 1 N NaOH. After neutralization, radioactivity was counted with a toluene scintillator.

#### *In Vivo* Antifungal Activities of FR209602 and FR209603 against *Candida albicans* in a Murine Infection Model

The *in vivo* anti-*Candida* activities were evaluated in a murine model of systemic infection using *Candida albicans* FP633. The inoculum was prepared from a three-day old culture of YM agar slant. ICR mice (female, four weeks old), were intravenously injected with  $1 \times 10^6$  cells of the yeast. Five mice were used in each group. Test compounds were dissolved in saline and administered subcutaneously one hour after challenge and once a day for three consecutive days. The ED<sub>50</sub> value was determined on

**Table 1** *In vitro* antifungal activities (MECs) of FR209602, FR209603, FR209604 and FR901379 by microbroth dilution method

	FR209602	FR209603	FR209604	FR901379
<i>Candida albicans</i> FP633	0.04	0.04	0.04	0.08
<i>Aspergillus fumigatus</i> FP1305	0.04	0.02	0.04	0.08
<i>Cryptococcus neoformans</i> YC203	>50	>50	>50	50

MEC ( $\mu\text{g/ml}$ ).**Table 2** Inhibitory activities of FR209602 and related compounds on 1,3- $\beta$ -glucan synthase prepared from *Candida albicans*

Compound	IC <sub>50</sub> ( $\mu\text{g/ml}$ )
FR209602	0.49
FR209603	0.64
FR209604	0.72
FR901379	0.77

the day when all control mice (vehicle only) died.

## Results

### *In Vitro* Activities of FR209602, FR209603 and FR209604

Table 1 shows the antifungal activities of FR20902, FR209603 and FR209604. They showed almost equivalent activities to FR901379 against *Candida albicans* FP633 and *A. fumigatus* FP1305. These compounds were also inactive against *Cryptococcus neoformans* as was FR901379. With all of these compounds, hyphal morphological changes were observed in *A. fumigatus*, i.e. hyphae abnormally grown, shortened, stunted and highly branched with bipolar or vesicular tips, swollen germ tubes and frequent balloon-like cells (data not shown). These morphological changes were characteristic of the inhibition of 1,3- $\beta$ -glucan synthesis. Table 2 shows the inhibitory activities of these compounds on  $\beta$ -1,3-glucan synthase derived from *Candida albicans* FP633. Significant inhibition of  $\beta$ -1,3-glucan synthase was observed in all of these compounds, which was consistent with the *in vitro* antifungal activities of these compounds.

### *In Vivo* Antifungal Activities of FR209602 and FR209603 against *Candida albicans* in a Murine Infection Model

*In vivo* antifungal activities of FR209602 and FR209603

**Table 3(a)** *In vivo* antifungal activities of FR209602 and FR901379 against *Candida albicans* in a murine infection model

	FR209602	FR901379
ED <sub>50</sub> (mg/kg)	2.0	1.2

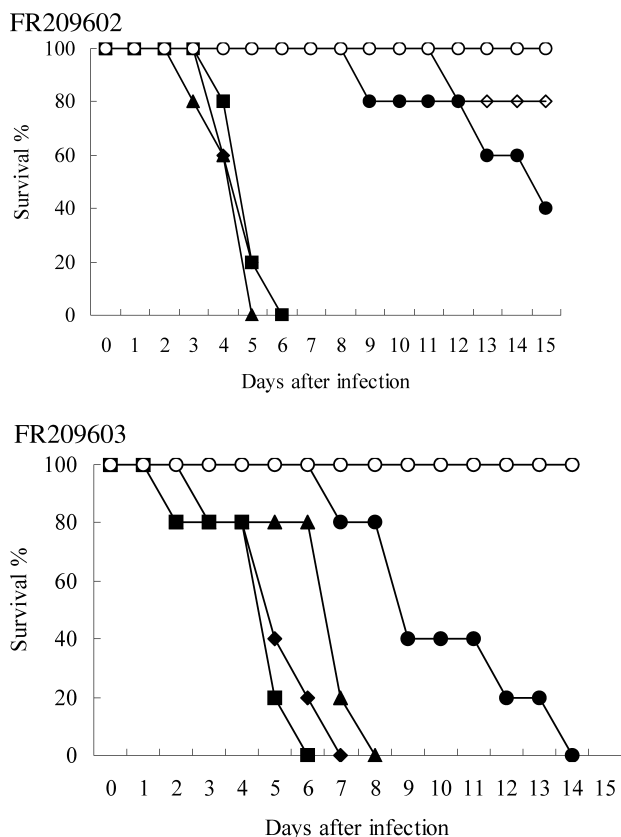
**Table 3(b)** *In vivo* antifungal activities of FR209603 and FR901379 against *Candida albicans* in a murine infection model

	FR209603	FR901379
ED <sub>50</sub> (mg/kg)	1.9	0.9

against *Candida albicans* in a murine infection model were evaluated, and compared to that of FR901379. Both FR209602 and FR209603 significantly prolonged the survival of infected mice by subcutaneously administration with ED<sub>50</sub> values of 2.0 mg/kg and 1.9 mg/kg, respectively (Table 3). These efficacies were almost equivalent to that of FR901379. Survival curves for the *in vivo* systemic infection model are shown in Fig. 2.

## Discussion

In this paper, we have presented *in vitro* and *in vivo* activities of novel antifungal lipopeptides, FR209602, FR209603 and FR209604, isolated from the fermentation broth of *Coleophoma crateriformis* No. 738. These compounds have potent antifungal activities against *Candida albicans* and *A. fumigatus*. The effects of these compounds on the morphology of *A. fumigatus* were characteristic of inhibition of 1,3- $\beta$ -glucan synthesis (data not shown). The IC<sub>50</sub> values of these compounds on *Candida albicans* 1,3- $\beta$ -glucan synthase were 0.49, 0.64 and 0.72  $\mu\text{g/ml}$ , respectively. Moreover, FR209602 and FR209603 were effective in a *Candida albicans* murine



**Fig. 2** Protective effect of FR209602 and FR209603 against systemic infection of *C. albicans* FP633.

◆, Control; ■, 0.32 mg/kg; ▲, 1.0 mg/kg; ●, 3.2 mg/kg; ◇, 10 mg/kg; ○, 32 mg/kg

infection model.

These compounds have similar structures to FR901379, the points of difference being the amino acid constituents of the cyclic peptide portion of their structures. It is known that amino acid composition affects both the antifungal spectrum as well as the chemical stability of lipopeptides. FR209602-4 could therefore provide a new direction for developing the next generation of antifungal agents.

## References

1. Wade JC. Treatment of fungal and other opportunistic infections in immunocompromised patients. *Leukemia* 11 (Suppl. 4): S38–S39 (1997)
2. Gallis HA, Drew RH, Pickard WW. Amphotericin B: 30 years of clinical experience. *Rev Infect Dis* 12: 308–329 (1990)
3. Carledge JD, Midgley J, Gazzard BG. Clinically significant azole cross-resistance in *Candida* isolates from HIV-positive patients with oral candidiasis. *AIDS* 11: 1839–1844 (1997)
4. Kanasaki R, Sakamoto K, Hashimoto M, Takase S, Tsurumi Y, Fujie A, Hino M, Hashimoto S, Hori Y. FR209602 and related compounds, novel antifungal lipopeptides from *Coleophoma crateriformis* No. 738. I. Taxonomy, fermentation, isolation and physico-chemical properties. *J Antibiot* 59: 137–144 (2006)
5. Liu J, Balasubramanian MK. 1,3-beta-Glucan synthase: a useful target for antifungal drugs. *Curr Drug Targets Infect Disord* 1: 159–169 (2001)
6. Tomishima M, Ohki H, Yamada A, Takasugi H, Maki K, Tawara S, Tanaka H. FK463, a novel water-soluble echinocandin lipopeptide: synthesis and antifungal activity. *J Antibiot* 52: 674–676 (1999)
7. Higashiyama Y, Kohno S. Micafungin: a therapeutic review. *Expert Rev Anti Infect Ther* 2: 345–355 (2004)
8. Jarvis B, Figgitt DP, Scott LJ. Micafungin. *Drugs* 64: 969–984 (2004)
9. Cornely OA, Schmitz K, Aisenbrey S. The first echinocandin: caspofungin. *Mycoses* 45: 56–60 (2002)
10. Iwamoto T, Fujie A, Sakamoto K, Tsurumi Y, Shigematsu N, Yamashita M, Hashimoto S, Okuhara M, Kohsaka M. WF11899A, B, and C, novel antifungal lipopeptides. I. Taxonomy, fermentation, isolation and physico-chemical properties. *J Antibiot* 47: 1084–1091 (1994)
11. Iwamoto T, Fujie A, Nitta K, Hashimoto S, Okuhara M, Kohsaka M. WF11899A, B, and C, novel antifungal lipopeptides. II. Biological properties. *J Antibiot* 47: 1092–1097 (1994)
12. Sawistowska-Schröder ET, Kerridge D, Perry H. Echinocandin inhibition of 1,3- $\beta$ -D-glucan synthase from *Candida albicans*. *FEBS Lett* 173: 134–138 (1984)